

An excerpt from the scientific article

Knowledge from human relevant cell, tissue and mathematics-based methods as key tools for understanding COVID-19

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CHAPTER 8.7.

Diamagnetism as an alternative or integrating cellular therapy for COVID-19

Naturally occurring electric fields are not only important for cell-surface interactions but are also pivotal for the normal development of the organism and its physiological functions. Selective control of cell function by applying specifically configured, weak, time-varying magnetic fields has added a therapeutic dimension to biology and medicine, e.g., diamagnetic therapies. The word “diamagnetic” originated from diamagnetism, which refers to the magnetic property of some materials which, subjected to a sufficiently intense magnetic field, receive a repulsive and distancing force with respect to the magnetic source. One of the most common diamagnetic materials is body water, and most proteins and ions are diamagnetic. Using diamagnetic therapy, there is a powerful effect on cellular features including specific cell membrane mechanisms reducing edema and inflammatory processes and the damaged tissue is quickly repaired with an immediate analgesic effect. Field parameters for therapeutic, pulsed electromagnetic field (PEMFs) were designed to induce voltages similar to those normally produced during dynamic mechanical deformation of e.g., in human body connective tissues. As a result, a wide variety of challenging musculoskeletal disorders have been treated successfully over the decennia. Patients with delayed union or non-union fractures have benefitted, from this surgically non-invasive method. Many of the athermal bioresponses, at the cellular and subcellular levels, have been identified and found appropriate to correct or modify the pathologic processes for which PEMFs have been used supported by double-blind trials. As understanding of underlying molecular and cellular mechanisms expand, specific requirements for field energetics are being defined and the range of therapeutic

broadened. These include nerve regeneration, wound healing, graft behavior, diabetes, and myocardial and cerebral ischemia (heart attack and stroke), among other conditions. [219]

The biophysical stimulation induced by Low Frequency – High-Intensity Pulsed Electromagnetic Fields (LF- HI- PEMF) – Diamagnetotherapy, has recently been demonstrated to be effective in the treatment of lung fibrosis associated to autoimmune diseases as well as to fibrosis in post COVID-19 pneumonia.

In both conditions that share a dysregulation of the immune system, diamagnetotherapy, ameliorated the functionality of respiratory muscles in treated patients, improved dyspnoea, oxygen saturation, indicative of a better performance of the lung. In these cases, a double effect in lung parenchyma and in respiratory muscles has been evaluated, also considering that in post-intensive care syndrome, patients report muscle weakness, impaired mobility and balance, joint stiffness while the associated neuropathy and myopathy are a consequence of SARS-CoV-2 infection. These clinical conditions worsen the morbidity and the mortality in COVID-19 patients. Diamagnetotherapy opens the possibility to treat symptoms in COVID-19 patients such as epilepsy, cognitive impairment, or motor imbalance, including the problems of the peripheral nervous system.

The rationale to employ diamagnetotherapy for medical problems is attributed to the intensity of the magnetic field that ensures the optimal and safe electric stimulation of the cell membranes. Moreover, the diamagnetic effect is also composed of a multi-variable spectrum of electromagnetic frequencies that selectively interact with the different electric state of the cell membrane in various situations. This phenomenon has been observed with electromyography following the stimulation of the motor cortex in healthy individuals. [220] This opens the possibility to treat symptoms in COVID-19 patients such as epilepsy, cognitive impairment, or motor imbalance, including the problems of the peripheral nervous system.

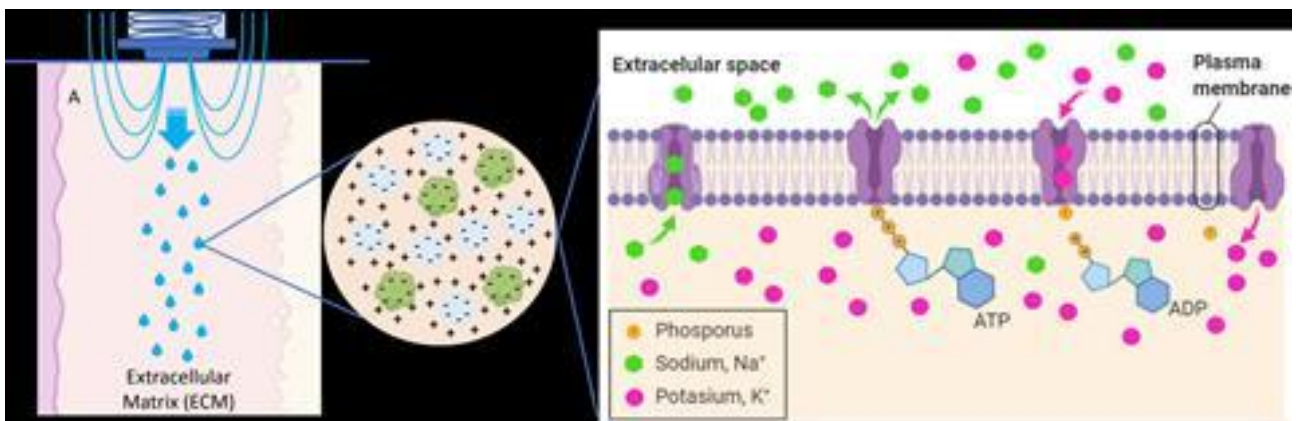
Diamagnetotherapy takes its name from a mechanical repulsive effect on diamagnetic substances e.g., water. 60% of the human adult body is water. Already in 1945 the % of water content in human body organs was described for an adult (Table 7) with an average weight of 70.5 kg. [221]

Table 7
Water content in human body organs/tissues.

Tissue	Water (%)
Lung	83.74
Striated Muscle	79.52
Kidney	79.47
Digestive tract	79.07
Spleen	78.69
Brain, spinal cord, nerve trunks	73.69
Hearth	73.69
Pancreas	73.08
Liver	71.46
Skin	64.86
Adipose tissue	50.09
Skeleton	31.81
Teeth	5
Liquid Tissues	93.33
Remaining Solid Tissues	70.40

The diamagnetic phenomenon moves liquids and solutes of the extracellular matrix (ECM) and of the intracellular environment. This, positively stimulates the metabolic activities of the treated tissues, reduce fibrosis and ameliorate peripheral oedema in limbs and the possibility to treat lymphatic imbalance related to the prolonged COVID-19 related immobility is a possible scenario.

Although different in terms of variability and strength of the physical features, HI magnetic fields have therapeutic biological effects in comparison to the more studied Low Intensity- PEMF with main regards to the anti-inflammatory, regenerative and trophic on the extracellular matrix (ECM). [222] The diversity consists in the interaction of HI-MF with a chain of diamagnetic nanoparticles of the cell membrane (ions, membrane receptor proteins, cholesterol, glycol, and phospholipids) and the intracellular cytosol. HI-MF modifies the hydrostatic pressure of the ECM and the transmembrane flux of ions reflecting on the electric potential of the cells. The phenomenon is enforced by the variability of the frequencies of the Electromagnetic Field (Figure 17).



Variable intensities of the magnetic field inhibit IL-6 and TNF- α expression at the gene level as well as IL-6, IL-1 β and TNF α expression at protein level in injured tissues. [223] This modulatory effect is confirmed in other studies that reveal the increase of the anti-inflammatory IL-10, observed in tendon cell cultures [224], and is also known for a strict relationship with the immune system in various, including autoimmune, pathologies. [225] Reference to COVID-19 induced immune dysregulation is consequential. Finally, PEMF preserve ECM integrity of the cultured embryonic cartilage explants by modulating the metabolism of proteoglycans without affecting their gross structural nature.

These experiences show that diamagnetism-based therapeutic interventions should be further explored as an alternative or integrating cellular therapy for COVID-19 for e.g., brain, immune system muscles and parenchymal organs.

The chronobiology of the inflammatory process relates to the of IL-1, IL-6 and TNF. [226] This physiological self-control and cellular self-defense of inflammation is shown in the response time of IL-10, and TGF-Beta and increase in the genetic expression of the interleukins involved in the inflammatory process. [227]

However, we know that the rapid and exaggerated increase in IL-6 in patients with COVID-19 presents a response that can lead to a chronic increase in IL-6 leading to a chronic activation of IL-17. This leads to a response in the activity of specific antibodies to tissues, cells, cell membranes and/or intracellular organelles, which may have

autoimmune or degenerative sequelae. However, the physiological response to the elevation of interleukin IL-6 and the subsequent elevation of IL-10 and TGF-beta [228] should be taken into account. As a consequence, we have an overexpression of these last two molecules, and the return to normality of TGF-beta is slow and usually occurs between 3-6 weeks after the acute pro-inflammatory event. However, the continuous activation of TGF-beta can cause changes in ECM [229], making it less soluble and evoking an increase in secondary fibrosis caused in the inflamed human body tissue areas. In the case of the post-COVID-19 patients, these tissues are identified by ACE receptors and through the stimulation given by chemotaxis and necrotaxis, these tissues change the inflammatory adaptation response of metalloproteinases. [230]

The ECM takes within its sol-gel regulation a management of remodeling and plasticity of the matrix. [231] This dynamic remodeling acts as a control in homeostasis, and protection of cell proliferation, migration and differentiation. As such, the ECM has a molecular filter control crucial for cell survival. [232] The ECM filter function is regulated by the electromagnetic and sterilization processes, which is maintained through the control of reactive oxygen species (ROS) and pH, the changes on the heparan sulfate chains thus stimulating the activation of metalloproteinases, capable of achieving changes in the sol-gel turn. [232], [233] The sol phase within the ECM has a protease activity, to initiate a reshaping of the matrix through the hydrolysis of proteoglycans. The maximum pH in this phase is 7.35, with sympathicotonic activation. The gel phase within the ECM has an anti-protease activity, performing a reconstruction and deposit of the matrix by means of the synthesis of matrix proteins. The pH peak in this phase is 7.45 so it is an alkaline phase with vagotonic activation. [234]

This regulation turn of the ECM is given by the activity of metalloproteinases (MMPs) that are 22 human proteolytic enzymes regulated by Th1 lymphocytes and therefore carry out degradation effects of the ECM. The regulation of these MMPs are made by the inhibitory tissue of metalloproteinases (TIMPs), which are 4 inhibitory proteins of MMP. These are regulated by Th2 lymphocytes and therefore carry out protein deposition in the ECM, this inhibition is carried out 1:1 from TIMPs to MMPs. [235], [236]

Therefore, inflammation as well as the increase in free radicals, increase the expression and secretion of the interleukins IL-1, IL-6 and TNF-alpha. This control in the degradation of matrix proteins is by means of the IL-4, IL-10 and TGF-beta1. This increases the deposit of glycoproteins and provides a control in physiological anti-inflammatory homeostasis. [237] Oxidative stress processes control cell aging. It is known that low levels of free radicals do not achieve the protection of cell survival that is needed and the high increase in oxidative stress causes a loss of oxidation reduction causing cell death and having low-grade chronic inflammation. [238]

ROS overexpression and pH reduction create pathological inflammation starting with an acute phase of inflammation like the one seen in COVID-19 in the acute and semi-acute phases, which enters into the sol phase as a degradative and inflammatory phase. At the same time, after the acute phase, a low-grade inflammation due to the increase in the expression and secretion of IL-6, causes as a regulatory control an increase in the secretion and expression of TGF-beta1, creating ECM rigidity due to an increase in the deposits of the proteoglycans giving the fibrotic phenotype. [239]

However, chronic alteration not only causes a rigidity of the ECM, it also creates an alteration of the intercellular junctions. In the case of inflammation caused by COVID-19, the mucosa represents alterations in the intercellular junctions of lung and receptors of the respiratory and oropharyngeal mucosa. In the processes of traditional inflammatory control, IL-10 activates the STAT3 receptors that decrease the expression of Claudin-2 and increase the expression of Claudin-4. TGF-beta activating the ERK and SMAD2 receptors, increasing the expression of Claudin-1, has been observed in autoimmune control processes in intestinal mucosa caused by Crohn's disease and treated with low interleukin dose and sequential kinetic activation. [240], [241], [242], [243]

An integrative treatment option with diamagnetic therapy is the implementation, penetration or implantation of therapeutic agents directly on the tissues under treatment. Due to the effect of diamagnetic repulsion movement, it is possible to introduce a product up to 8-10 cm deep (Figure 18).

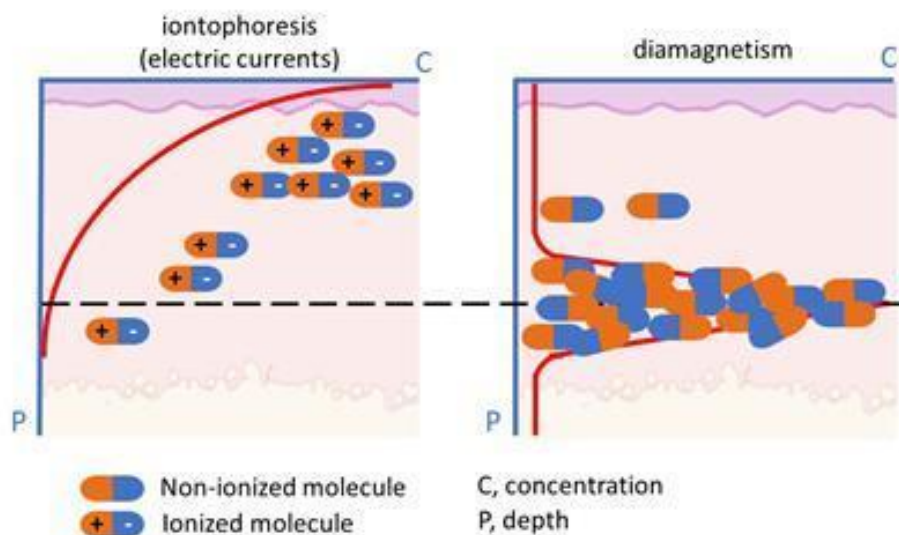


Figure 18

Distribution curve of pharmacological substances (physical model) expressed as concentration-depth ratio (PC) induced respectively by electric currents or High Intensity-PEMF Iontophoresis (electric currents). The delivery of the therapeutic agent decreases with the distance and depends on the ionization of the molecules. B) Homogeneous and deeper distribution of hydro-soluble (diamagnetic) therapeutic agent as a result of the impulse originating from the High Intensity of Magnetic Field. The ionization of the molecule is not necessary.

Therapeutic agents with the greatest possibility of transport are interleukins, hormones, neuropeptides or growth factors at low dose, which have already shown adequate control of the processes of low-grade chronic inflammation which causes COVID-19 inflammatory, autoimmune or degenerative sequelae, that is observed in patients between 2-3 weeks after the acute phase and up to 3-4 months. [244]

References

1. Gandhi, R.T., J.B. Lynch, and C. Del Rio, *Mild or Moderate Covid-19*. N Engl J Med, 2020. **383** (38): p. 1757-1766.
2. Fajgenbaum, D.C. and C.H. June, *Cytokine Storm*. N Engl J Med, 2020. **383** (23): p. 2255-2273.
3. WHO. *Coronavirus disease (COVID-19) outbreak situation*. Available from: <https://www.who.int/>.
4. Walsh, K.A., et al., *SARS-CoV-2 detection, viral load and infectivity over the course of an infection*. J Infect, 2020. **81** (3): p. 357-371.
5. Bermejo-Martin, J.F., González-Rivera, M., Almansa, R., and e. al., *Viral RNA load in plasma is associated with critical illness and a dysregulated host response in COVID-19*. Crit Care, 2020. **24**: p. 691.
6. European Centre for Disease Prevention and Control, A.a.o.t.E.U. *Diagnostic testing and screening for SARS-CoV-2*. 2020; Available from: <https://www.ecdc.europa.eu/en/covid-19/latest-evidence/diagnostic-testing>.
7. Joint Research Centre, J., *The JRC releases new reference materials for the quality control of SARS-CoV-2 antibody tests*. 2020.
8. Wang, S., et al., *Modeling the viral dynamics of SARS-CoV-2 infection*. Mathematical Biosciences, 2020. **328**.
9. Chan, J.F., et al., *Genomic characterization of the 2019 novel human-pathogenic coronavirus isolated from a patient with atypical pneumonia after visiting Wuhan*. Emerg Microbes Infect, 2020. **9** (1): p. 221-236.
10. Kadam, S.B., et al., *SARS-CoV-2, the pandemic coronavirus: Molecular and structural insights*. J Basic Microbiol, 2021.
11. Jing, Y., et al., *Potential influence of COVID-19/ACE2 on the female reproductive system*. Mol Hum Reprod, 2020. **26** (6): p. 367-373.
12. Pillaiyar, T., et al., *The recent outbreaks of human coronaviruses: A medicinal chemistry perspective*. Med Res Rev, 2020.
13. Battagello, D.S., et al., *Unpuzzling COVID-19: tissue-related signaling pathways associated with SARS-CoV-2 infection and transmission*. Clin Sci (Lond), 2020. **134** (16): p. 2137-2160.
14. Li, H., et al., *Transmission Routes Analysis of SARS-CoV-2: A Systematic Review and Case Report*. Front Cell Dev Biol, 2020. **8**: p. 618.
15. Mohseni, A.H., et al., *Body fluids may contribute to human-to-human transmission of severe acute respiratory syndrome coronavirus 2: evidence and practical experience*. Chin Med, 2020. **15**: p. 58.
16. Busquet, F., et al., *Harnessing the power of novel animal-free test methods for the development of COVID-19 drugs and vaccines*. Arch Toxicol, 2020. **94** (6): p. 2263-2272.
17. *Animals used for scientific purposes, Replacement, Reduction and Refinement – the “Three Rs”*. Available from: https://ec.europa.eu/environment/chemicals/lab_animals/3r/alternative_en.htm.
18. Agency, E.C., *New Approach Methodologies in Regulatory Science Proceedings of a scientific workshop*. 2016.
19. Punt, A., et al., *New approach methodologies (NAMs) for human-relevant biokinetics predictions. Meeting the paradigm shift in toxicology towards an animal-free chemical risk assessment*. ALTEX, 2020. **37** (4): p. 607-622.
20. Khoury, D.S., et al., *Measuring immunity to SARS-CoV-2 infection: comparing assays and animal models*. Nat Rev Immunol, 2020. **20** (12): p. 727-738.
21. Takayama, K., *In Vitro and Animal Models for SARS-CoV-2 research*. Trends Pharmacol Sci, 2020. **41** (8): p. 513-517.
22. Munoz-Fontela, C., et al., *Animal models for COVID-19*. Nature, 2020. **586** (7830): p. 509-515.
23. Coecke, S., et al., *Guidance on Good Cell Culture Practice, A Report of the Second ECVAM Task Force on Good Cell Culture Practice in Cell Technology for Cell Products*. R. Smith, Editor. 2007. p. 313-315.
24. OECD, *Guidance Document on Good In Vitro Method Practices (GIVIMP), OECD Series on Testing and Assessment*. Vol. 286. 2018: OECD Publishing.
25. Aydin, A., et al., *Combating COVID-19 with tissue engineering: a review*. Emergent Mater, 2020: p. 1-21.
26. Pizzorno, A., et al., *Characterization and Treatment of SARS-CoV-2 in Nasal and Bronchial Human Airway Epithelia*. Cell Rep Med, 2020. **1** (4): p. 100059.
27. Bullen, C.K., et al., *Infectability of human BrainSphere neurons suggests neurotropism of SARS-CoV-2*. ALTEX, 2020. **37** (4): p. 665-671.
28. de Melo, B.A.G., et al., *3D culture models to study SARS-CoV-2 infectivity and antiviral candidates: From spheroids to bioprinting*. Biomedical Journal, 2020.
29. Hao, S., et al., *Long-Term Modeling of SARS-CoV-2 Infection of In Vitro Cultured Polarized Human Airway Epithelium*. mBio, 2020. **11** (6).
30. Monteil, V., et al., *Inhibition of SARS-CoV-2 Infections in Engineered Human Tissues Using Clinical-Grade Soluble Human ACE2*. Cell, 2020. **181** (4): p. 905-913 e7.
31. Chakraborty, J., et al., *Bioengineered in Vitro Tissue Models to Study SARS-CoV-2 Pathogenesis and Therapeutic Validation*. ACS Biomater Sci Eng, 2020. **6** (12): p. 6540-6555.
32. Simoneau, C.R. and M. Ott, *Modeling Multi-organ Infection by SARS-CoV-2 Using Stem Cell Technology*. Cell Stem Cell, 2020. **27** (6): p. 859-868.
33. Assaf, D., et al., *Utilization of machine-learning models to accurately predict the risk for critical COVID-19*. Intern Emerg Med, 2020. **15** (8): p. 1435-1443.
34. Rangel, H.R., et al., *SARS-CoV-2 host tropism: An in silico analysis of the main cellular factors*. Virus Research, 2020. **289**: p. 198154.
35. Hussain, M., et al., *Molecular docking between human TMPRSS2 and SARS-CoV-2 spike protein: conformation and intermolecular interactions*. AIMS Microbiol, 2020. **6** (3): p. 350-360.
36. Ziegler, C.G.K., et al., *SARS-CoV-2 Receptor ACE2 Is an Interferon-Stimulated Gene in Human Airway Epithelial Cells and Is Detected in Specific Cell Subsets across Tissues*. Cell, 2020. **181** (5): p. 1016-1035 e19.
37. Doran, K.S., et al., *Concepts and mechanisms: Crossing host barriers*. Cold Spring Harbor Perspectives in Medicine, 2013. **3** (7).
38. Vielle, N.J., et al., *The human upper respiratory tract epithelium is susceptible to flaviviruses*. Frontiers in Microbiology, 2019. **10** (MAR).
39. Invernizzi, R., C.M. Lloyd, and P.L. Molyneaux, *Respiratory microbiome and epithelial interactions shape immunity in the lungs*. Immunology, 2020. **160** (2): p. 171-182.
40. Sharma, L. and A. Riva, *Intestinal barrier function in health and disease—any role of sars-cov-2?* Microorganisms, 2020. **8** (11): p. 1-27.
41. Lee, J.J., et al., *Relative abundance of sars-cov-2 entry genes in the enterocytes of the lower gastrointestinal tract*. Genes, 2020. **11** (6): p. 1-9.
42. Chu, H., et al., *Comparative tropism, replication kinetics, and cell damage profiling of SARS-CoV-2 and SARS-CoV with implications for clinical manifestations, transmissibility, and laboratory studies of COVID-19: an observational study*. The Lancet Microbe, 2020. **1** (1): p. e14-e23.
43. Lamers, M.M., et al., *SARS-CoV-2 productively infects human gut enterocytes*. Science, 2020. **369** (6499): p. 50-54.
44. Imura, Y., et al., *A microfluidic system to evaluate intestinal absorption*. Anal Sci, 2009. **25** (12): p. 1403-7.
45. Chi, M., et al., *A microfluidic cell culture device (muFCCD) to culture epithelial cells with physiological and morphological properties that mimic those of the human intestine*. Biomed Microdevices, 2015. **17** (3): p. 9966.
46. Invernizzi, A., et al., *Retinal findings in patients with COVID-19: Results from the SERPICO-19 study*. EClinicalMedicine, 2020. **27**: p. 100550.
47. Hirose, R., et al., *Survival of SARS-CoV-2 and influenza virus on the human skin: Importance of hand hygiene in COVID-19*. Clin Infect Dis, 2020.
48. Mao, L., et al., *Neurologic Manifestations of Hospitalized Patients With Coronavirus Disease 2019 in Wuhan, China*. JAMA Neurol, 2020. **77** (6): p. 683-690.
49. Xu, J. and E. Lazartigues, *Expression of ACE2 in Human Neurons Supports the Neuro-Invasive Potential of COVID-19 Virus*. Cell Mol Neurobiol, 2020.

50. Helms, H.C., et al., *In vitro models of the blood-brain barrier: An overview of commonly used brain endothelial cell culture models and guidelines for their use*. J Cereb Blood Flow Metab, 2016. **36** (5): p. 862-90.
51. Prieto, P., et al., *Blood-brain barrier in vitro models and their application in toxicology. The report and recommendations of ECVAM Workshop 49*. Altern Lab Anim, 2004. **32** (1): p. 37-50.
52. Buzhdygan, T.P., et al., *The SARS-CoV-2 spike protein alters barrier function in 2D static and 3D microfluidic in vitro models of the human blood-brain barrier*. bioRxiv, 2020.
53. Rees, S., R. Harding, and D. Walker, *The biological basis of injury and neuroprotection in the fetal and neonatal brain*. Int J Dev Neurosci, 2011. **29** (6): p. 551-8.
54. Leon-Juarez, M., et al., *Cellular and molecular mechanisms of viral infection in the human placenta*. Pathog Dis, 2017. **75** (7).
55. Alzamora, M.C., et al., *Severe COVID-19 during Pregnancy and Possible Vertical Transmission*. Am J Perinatol, 2020. **37** (8): p. 861-865.
56. Wong, M.K., et al., *Establishment of an in vitro placental barrier model cultured under physiologically relevant oxygen levels*. Mol Hum Reprod, 2020. **26** (5): p. 353-365.
57. Nishiguchi, A., et al., *In vitro placenta barrier model using primary human trophoblasts, underlying connective tissue and vascular endothelium*. Biomaterials, 2019. **192**: p. 140-148.
58. Blundell, C., *A microphysiological model of the human placental barrier*. Lab Chip, 2016. **16** (16): p. 3065-73.
59. Mital, P., B.T. Hinton, and J.M. Dufour, *The blood-testis and blood-epididymis barriers are more than just their tight junctions*. Biol Reprod, 2011. **84** (5): p. 851-8.
60. Cheng, C.Y. and D.D. Mruk, *The blood-testis barrier and its implications for male contraception*. Pharmacol Rev, 2012. **64** (1): p. 16-64.
61. Verma, S., S. Saksena, and H. Sadri-Ardekani, *ACE2 receptor expression in testes: implications in coronavirus disease 2019 pathogenesis*. Biol Reprod, 2020. **103** (3): p. 449-451.
62. Wambier, C.G. and A. Goren, *Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection is likely to be androgen mediated*. J Am Acad Dermatol, 2020. **83** (1): p. 308-309.
63. Mruk, D.D. and C.Y. Cheng, *An in vitro system to study Sertoli cell blood-testis barrier dynamics*. Methods Mol Biol, 2011. **763**: p. 237-52.
64. Byers, S.W., et al., *Growth and characterization of polarized monolayers of epididymal epithelial cells and Sertoli cells in dual environment culture chambers*. J Androl, 1986. **7** (1): p. 59-68.
65. Yamada, K.M. and E. Cukierman, *Modeling tissue morphogenesis and cancer in 3D*. Cell, 2007. **130** (4): p. 601-10.
66. Steinberger, A., E. Steinberger, and W.H. Perloff, *Mammalian testes in organ culture*. Experimental Cell Research, 1964. **36** (1): p. 19-27.
67. Sakib, S., et al., *Three-dimensional testicular organoids as novel in vitro models of testicular biology and toxicology*. Environ Epigenet, 2019. **5** (3): p. dvz011.
68. Alves-Lopes, J.P. and J.B. Stukenborg, *Testicular organoids: a new model to study the testicular microenvironment in vitro?* Hum Reprod Update, 2018. **24** (2): p. 176-191.
69. Lancaster, M.A. and J.A. Knoblich, *Organogenesis in a dish: modeling development and disease using organoid technologies*. Science, 2014. **345** (6194): p. 1247125.
70. Millet, J.K., J.A. Jaimes, and G.R. Whittaker, *Molecular diversity of coronavirus host cell entry receptors*. FEMS Microbiol Rev, 2020.
71. Huertas, A.M., D.; Savale, L.; Pichon, J.; Tu, Y.; Parent, F.; Guignabert, C.; Humbert, M., *Endothelial cell dysfunction: a major player in SARS-CoV-2 infection (COVID-19)?* European Respiratory Journal, 2020. **56**: p. 2001634.
72. Essahib, W., et al., *SARS-CoV-2 host receptors ACE2 and CD147 (BSG) are present on human oocytes and blastocysts*. J Assist Reprod Genet, 2020. **37** (11): p. 2657-2660.
73. Amraie, R., et al., *CD209/L-SIGN and CD209/DC-SIGN act as receptors for SARS-CoV-2 and are differentially expressed in lung and kidney epithelial and endothelial cells*. bioRxiv, 2020.
74. Naffah-Mazzacoratti Mda, G., et al., *What have we learned about the kallikrein-kinin and renin-angiotensin systems in neurological disorders?* World J Biol Chem, 2014. **5** (2): p. 130-40.
75. Carvalho, P.R., P. Sirois, and P.D. Fernandes, *The role of kallikrein-kinin and renin-angiotensin systems in COVID-19 infection*. Peptides, 2020. **135**: p. 170428.
76. Zamorano Cuervo, N. and N. Grandvaux, *ACE2: Evidence of role as entry receptor for SARS-CoV-2 and implications in comorbidities*. Elife, 2020. **9**.
77. Tang, H., et al., *Thoughts on detecting tissue distribution of potential COVID-19 receptors*. Future Virology, 2020. **15** (8): p. 489-496.
78. Matusiak, M. and C.M. Schurch, *Expression of SARS-CoV-2 entry receptors in the respiratory tract of healthy individuals, smokers and asthmatics*. Respir Res, 2020. **21** (3): p. 252.
79. Li, M.Y., et al., *Expression of the SARS-CoV-2 cell receptor gene ACE2 in a wide variety of human tissues*. Infect Dis Poverty, 2020. **9** (1): p. 45.
80. Vaajänen, A., et al., *The expression of Mas-receptor of the renin-angiotensin system in the human eye*. Graefes Arch Clin Exp Ophthalmol, 2015. **253** (7): p. 1053-9.
81. Tseng, Y.H., R.C. Yang, and T.S. Lu, *Two hits to the renin-angiotensin system may play a key role in severe COVID-19*. Kaohsiung J Med Sci, 2020. **36** (6): p. 389-392.
82. Jackson, L., et al., *Within the Brain: The Renin Angiotensin System*. Int J Mol Sci, 2018. **19** (3).
83. Gupta, A., et al., *Extrapulmonary manifestations of COVID-19*. Nat Med, 2020. **26** (7): p. 1017-1032.
84. Huang, C., et al., *Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China*. The Lancet, 2020. **395** (10223): p. 497-506.
85. Zhou, F., et al., *Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study*. The Lancet, 2020. **395** (10229): p. 1054-1062.
86. Yang, X., et al., *Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: a single-centered, retrospective, observational study*. The Lancet Respiratory Medicine, 2020. **8** (5): p. 475-481.
87. Chen, G., et al., *Clinical and immunological features of severe and moderate coronavirus disease 2019*. J Clin Invest, 2020. **130** (5): p. 2620-2629.
88. Recalcati, S., *Cutaneous manifestations in COVID-19: a first perspective*. J Eur Acad Dermatol Venerol, 2020. **34** (5): p. e212-e213.
89. Wan, J., et al., *Digestive symptoms and liver injury in patients with coronavirus disease 2019 (COVID-19): A systematic review with meta-analysis*. JGH Open, 2020. **4** (6): p. 1047-1058.
90. Goncalves, L.F., et al., *Smell and taste alterations in COVID-19 patients: a systematic review*. Rev Assoc Med Bras (1992), 2020. **66** (11): p. 1602-1608.
91. da Rosa Mesquita, R., et al., *Clinical manifestations of COVID-19 in the general population: systematic review*. Wien Klin Wochenschr, 2020.
92. MadaniNeishaboori, A., et al., *Central Nervous System Complications in COVID-19 Patients; a Systematic Review and Meta-Analysis based on Current Evidence*. Arch Acad Emerg Med, 2020. **8** (1): p. e62.
93. Islam, M.A., et al., *Prevalence of Headache in Patients With Coronavirus Disease 2019 (COVID-19): A Systematic Review and Meta-Analysis of 14,275 Patients*. Front Neurol, 2020. **11**: p. 562634.
94. Guan, W., et al., *Clinical characteristics of coronavirus disease 2019 in China*. New England Journal of Medicine, 2020. **382** (18): p. 1708-1720.
95. Pascarella, G., et al., *COVID-19 diagnosis and management: a comprehensive review*. Journal of Internal Medicine, 2020. **288** (2): p. 192-206.
96. Lee, I.T., et al., *ACE2 localizes to the respiratory cilia and is not increased by ACE inhibitors or ARBs*. Nat Commun, 2020. **11** (1): p. 5453.
97. Katsura, H., et al., *Human Lung Stem Cell-Based Alveolospheres Provide Insights into SARS-CoV-2-Mediated Interferon Responses and Pneumocyte Dysfunction*. Cell Stem Cell, 2020. **27** (6): p. 890-904 e8.
98. Mason, R.J., *Pathogenesis of COVID-19 from a cell biology perspective*. European Respiratory Journal, 2020. **55** (4).
99. Mann, R., et al., *Clinical Characteristics, Diagnosis, and Treatment of Major Coronavirus Outbreaks*. Front Med (Lausanne), 2020. **7**: p. 581521.

100. Tahvildari, A., et al., *Clinical features, Diagnosis, and Treatment of COVID-19: A systematic review of case reports and case series* medRxiv preprint, 2020.
101. Kotta, S., et al., *Combating the Pandemic COVID-19: Clinical Trials, Therapies and Perspectives*. Front Mol Biosci, 2020. 7: p. 606393.
102. Santacroce, L., et al., *The human coronaviruses (HCoVs) and the molecular mechanisms of SARS-CoV-2 infection*. J Mol Med (Berl), 2020.
103. Machhi, J., et al., *The Natural History, Pathobiology, and Clinical Manifestations of SARS-CoV-2 Infections*. J Neuroimmune Pharmacol, 2020. 15 (3): p. 359-386.
104. Leisman, D.E., et al., *Cytokine elevation in severe and critical COVID-19: a rapid systematic review, meta-analysis, and comparison with other inflammatory syndromes*. The Lancet Respiratory Medicine, 2020. 8 (12): p. 1233-1244.
105. Mehta, P., et al., *COVID-19: consider cytokine storm syndromes and immunosuppression*. The Lancet, 2020. 395 (10229): p. 1033-1034.
106. Wigen, J., et al., *Converging pathways in pulmonary fibrosis and Covid-19 - The fibrotic link to disease severity*. Respir Med X, 2020. 2: p. 100023.
107. Singh, V., *Textbook of Anatomy: Abdomen and Lower Limb*. 2018. p. 24.
108. Puelles, V.G., et al., *Multifocal and Renal Tropism of SARS-CoV-2*. N Engl J Med, 2020. 383 (6): p. 590-592.
109. Tavazzi, G., et al., *Myocardial localization of coronavirus in COVID-19 cardiogenic shock*. Eur J Heart Fail, 2020. 22 (5): p. 911-915.
110. Wichmann, D., et al., *Autopsy Findings and Venous Thromboembolism in Patients With COVID-19: A Prospective Cohort Study*. Ann Intern Med, 2020. 173 (4): p. 268-277.
111. Sala, S., et al., *Acute myocarditis presenting as a reverse Tako-Tsubo syndrome in a patient with SARS-CoV-2 respiratory infection*. Eur Heart J, 2020. 41 (19): p. 1861-1862.
112. Xu, Z., et al., *Pathological findings of COVID-19 associated with acute respiratory distress syndrome*. The Lancet Respiratory Medicine, 2020. 8 (4): p. 420-422.
113. Varga, Z., et al., *Endothelial cell infection and endotheliitis in COVID-19*. The Lancet, 2020. 395 (10234): p. 1417-1418.
114. Li, H., et al., *SARS-CoV-2 and viral sepsis: observations and hypotheses*. The Lancet, 2020. 395 (10235): p. 1517-1520.
115. Bailey, A.L., et al., *SARS-CoV-2 Infects Human Engineered Heart Tissues and Models COVID-19 Myocarditis*. bioRxiv, 2020.
116. Mao, R., et al., *Manifestations and prognosis of gastrointestinal and liver involvement in patients with COVID-19: a systematic review and meta-analysis*. The Lancet Gastroenterology & Hepatology, 2020. 5 (7): p. 667-678.
117. Zhao, Y., et al., *COVID-19 and gastrointestinal symptoms*. Br J Surg, 2020. 107 (10): p. e382-e383.
118. Zhang, C., L. Shi, and F.-S. Wang, *Liver injury in COVID-19: management and challenges*. The Lancet Gastroenterology & Hepatology, 2020. 5 (5): p. 428-430.
119. Fan, Z., et al., *Clinical Features of COVID-19-Related Liver Functional Abnormality*. Clinical Gastroenterology and Hepatology, 2020. 18 (7): p. 1561-1566.
120. Lamers, M.M., et al., *SARS-CoV-2 productively infects human gut enterocytes*. Science, 2020. 369 (6499): p. 50-54.
121. Zhang, H., et al., *Digestive system is a potential route of COVID-19: an analysis of single-cell coexpression pattern of key proteins in viral entry process*. Gut, 2020. 69 (6): p. 1010-1018.
122. Wolfel, R., et al., *Virological assessment of hospitalized patients with COVID-2019*. Nature, 2020. 581 (7809): p. 465-469.
123. Xiao, F., et al., *Evidence for Gastrointestinal Infection of SARS-CoV-2*. Gastroenterology, 2020. 158 (6): p. 1831-1833 e3.
124. Guo, Y., et al., *Modeling SARS-CoV-2 infection in vitro with a human intestine-on-chip device*. BioRxiv, 2020.
125. Zou, X., et al., *Single-cell RNA-seq data analysis on the receptor ACE2 expression reveals the potential risk of different human organs vulnerable to 2019-nCoV infection*. Front Med (Lausanne), 2020. 14 (2): p. 185-192.
126. Cabibbo, G., et al., *SARS-CoV-2 infection in patients with a normal or abnormal liver*. J Viral Hepat, 2020.
127. Wang, Y., et al., *SARS-CoV-2 infection of the liver directly contributes to hepatic impairment in patients with COVID-19*. J Hepatol, 2020. 73 (4): p. 807-816.
128. Zhao, B., et al., *Recapitulation of SARS-CoV-2 infection and cholangiocyte damage with human liver ductal organoids*. Protein Cell, 2020. 11 (10): p. 771-775.
129. F.; Zhang T.; Kim T. W.; Harschnitz O.; Redmond D.; Houghton S.; Liu C.; Naji A.; Ciceri G.; Guttikonda S.; Bram Y.; Nguyen D.-H. T.; Cioffi M.; Chandar V.; Hoagland D. A.; Huang Y.; Xiang J.; Wang H.; Lyden D.; Borczuk A.; Chen H. J.; Studer L.; Pan F. C.; Ho D. D.; tenOever B. R.; Evans T.; Schwartz R. E.; Chen S., Y.L.H.Y.N.-P.B.E.G.V.W.P.D.X.T.X.Z.J.Z.J., *A Human Pluripotent Stem Cell-Based Platform to Study SARS-CoV-2 Tropism and Model Virus Infection in Human Cells and Organoids* Cell Stem Cell, 2020. 27: p. 125.
130. Su, H., et al., *Renal histopathological analysis of 26 postmortem findings of patients with COVID-19 in China*. Kidney International, 2020. 98 (1): p. 219-227.
131. Pan, X.W., et al., *Identification of a potential mechanism of acute kidney injury during the COVID-19 outbreak: a study based on single-cell transcriptome analysis*. Intensive Care Med, 2020. 46 (6): p. 1114-1116.
132. Iwasaki, A. and P.S. Pillai, *Innate immunity to influenza virus infection*. Nat Rev Immunol, 2014. 14 (5): p. 315-28.
133. Allison, S.J., *SARS-CoV-2 infection of kidney organoids prevented with soluble human ACE2*. Nat Rev Nephrol, 2020. 16 (6): p. 316.
134. Ryan, H. and C.S. Simmons, *Potential Applications of Microfluidics to Acute Kidney Injury Associated with Viral Infection*. Cell Mol Bioeng, 2020. 13 (4): p. 1-7.
135. DG, S. *The Long-Term Health impacts of Being Infected With the Coronavirus*. 2020; Available from: <https://elemental.medium.com/the-long-term-health-impacts-of-being-infected-with-the-coronavirus-d3a03f3cb6e8>.
136. Ferrario, C.M., *ACE2: more of Ang-(1-7) or less Ang II?* Curr Opin Nephrol Hypertens, 2011. 20 (1): p. 1-6.
137. Ashary, N., et al., *Single-Cell RNA-seq Identifies Cell Subsets in Human Placenta That Highly Expresses Factors Driving Pathogenesis of SARS-CoV-2*. Frontiers in Cell and Developmental Biology, 2020. 8.
138. Kubota, T. and N. Kuroda, *Exacerbation of neurological symptoms and COVID-19 severity in patients with preexisting neurological disorders and COVID-19: A systematic review*. Clin Neurol Neurosurg, 2020: p. 106349.
139. Achar, A. and C. Ghosh, *COVID-19-Associated Neurological Disorders: The Potential Route of CNS Invasion and Blood-Brain Relevance*. Cells, 2020. 9 (11).
140. Zhang, Y., et al., *Mechanisms involved in the development of thrombocytopenia in patients with COVID-19*. Thromb Res, 2020. 193: p. 110-115.
141. Chou, D.B., et al., *On-chip recapitulation of clinical bone marrow toxicities and patient-specific pathophysiology*. Nat Biomed Eng, 2020. 4 (4): p. 394-406.
142. Disser, N.P., et al., *Musculoskeletal Consequences of COVID-19*. J Bone Joint Surg Am, 2020. 102 (14): p. 1197-1204.
143. Grant, M.C., et al., *The prevalence of symptoms in 24,410 adults infected by the novel coronavirus (SARS-CoV-2; COVID-19): A systematic review and meta-analysis of 148 studies from 9 countries*. PLoS One, 2020. 15 (6): p. e0234765.
144. Parisi, S., et al., *Viral arthritis and COVID-19*. The Lancet Rheumatology, 2020. 2 (11): p. e655-e657.
145. Flaherty, K.R., et al., *Idiopathic pulmonary fibrosis: prognostic value of changes in physiology and six-minute-walk test*. Am J Respir Crit Care Med, 2006. 174 (7): p. 803-9.
146. Gupta, S., R.J.S. Dhillon, and S. Hasni, *Sarcopenia: A Rheumatic Disease?* Rheum Dis Clin North Am, 2018. 44 (3): p. 393-404.
147. Morley, J.E., K. Kalantar-Zadeh, and S.D. Anker, *COVID-19: a major cause of cachexia and sarcopenia?* J Cachexia Sarcopenia Muscle, 2020. 11 (4): p. 863-865.
148. Kirk, B., J. Zanker, and G. Duque, *Osteosarcopenia: epidemiology, diagnosis, and treatment-facts and numbers*. J Cachexia Sarcopenia Muscle, 2020. 11 (3): p. 609-618.
149. Dhillon, R.J. and S. Hasni, *Pathogenesis and Management of Sarcopenia*. Clin Geriatr Med, 2017. 33 (1): p. 17-26.
150. Paliwal, V.K., et al., *Neuromuscular presentations in patients with COVID-19*. Neurol Sci, 2020. 41 (11): p. 3039-3056.

151. Lad, H., et al., *Intensive Care Unit-Acquired Weakness: Not just Another Muscle Atrophiying Condition*. Int J Mol Sci, 2020. **21** (21): p. 7840.
152. Drenovska, K., E. Schmidt, and S. Vassileva, *Covid-19 pandemic and the skin*. Int J Dermatol, 2020.
153. Daneshgaran, G., D.P. Dubin, and D.J. Gould, *Cutaneous Manifestations of COVID-19: An Evidence-Based Review*. Am J Clin Dermatol, 2020. **21** (5): p. 627-639.
154. Conforti, C., et al., *Cutaneous Manifestations in Confirmed COVID-19 Patients: A Systematic Review*. Biology (Basel), 2020. **9** (12).
155. Hubiche, T., et al., *Clinical, Laboratory, and Interferon-Alpha Response Characteristics of Patients With Chilblain-like Lesions During the COVID-19 Pandemic*. JAMA Dermatol, 2020.
156. Colonna, C., et al., *Chilblain-like lesions in children following suspected COVID-19 infection*. Pediatr Dermatol, 2020. **37** (3): p. 437-440.
157. Al-Benna, S., *Gene Expression of Angiotensin-Converting Enzyme 2 Receptor in Skin and the Implications for COVID-19*. Adv Skin Wound Care, 2021. **34** (1): p. 31-35.
158. Matei, A.E., et al., *Vascularised human skin equivalents as a novel in vitro model of skin fibrosis and platform for testing of antifibrotic drugs*. Ann Rheum Dis, 2019. **78** (12): p. 1686-1692.
159. Al Heialy, S., et al., *Regulation of Angiotensin- Converting Enzyme 2 in Obesity: Implications for COVID-19*. Front Physiol, 2020. **11**: p. 555039.
160. Pasquarelli-do-Nascimento, G., et al., *Hypercoagulopathy and Adipose Tissue Exacerbated Inflammation May Explain Higher Mortality in COVID-19 Patients With Obesity*. Front Endocrinol (Lausanne), 2020. **11**: p. 530.
161. Olaniyan, O.T., et al., *Testis and blood-testis barrier in Covid-19 infestation: role of angiotensin-converting enzyme 2 in male infertility*. J Basic Clin Physiol Pharmacol, 2020. **31** (6).
162. Ponten, F., K. Jirstrom, and M. Uhlen, *The Human Protein Atlas—a tool for pathology*. J Pathol, 2008. **216** (4): p. 387-93.
163. *The Human Proteome Atlas*. Available from: <https://www.proteinatlas.org/ENSG00000130234-ACE2/tissue>.
164. Yang, M., et al., *Pathological Findings in the Testes of COVID-19 Patients: Clinical Implications*. Eur Urol Focus, 2020. **6** (5): p. 1124-1129.
165. Brannen, K.C., et al., *Alternative Models of Developmental and Reproductive Toxicity in Pharmaceutical Risk Assessment and the 3Rs*. ILAR J, 2016. **57** (2): p. 144-156.
166. Zheng, K., et al., *COVID-19 and the bone: underestimated to consider*. Eur Rev Med Pharmacol Sci, 2020. **24** (20): p. 10316-10318.
167. Lespasio, M.J., N. Sodhi, and M.A. Mont, *Osteonecrosis of the Hip: A Primer*. Perm J, 2019. **23**.
168. Cuetara, B.L., et al., *Cloning and characterization of osteoclast precursors from the RAW264.7 cell line*. In Vitro Cell Dev Biol Anim, 2006. **42** (7): p. 182-8.
169. Li, J., et al., *COVID-19 infection may cause ketosis and ketoacidosis*. Diabetes Obes Metab, 2020. **22** (10): p. 1935-1941.
170. Liu, F., et al., *Highly ACE2 Expression in Pancreas May Cause Pancreas Damage After SARS-CoV-2 Infection*. medRxiv preprint, 2020.
171. Scappaticcio, L., et al., *Impact of COVID-19 on the thyroid gland: an update*. Rev Endocr Metab Disord, 2020.
172. Valencia, I., et al., *DPP4 and ACE2 in Diabetes and COVID-19: Therapeutic Targets for Cardiovascular Complications?* Front Pharmacol, 2020. **11**: p. 1161.
173. Roncati, L. and B. Lusenti, *The <moonlighting protein>> able to explain the Th1 immune lockdown in severe COVID-19*. Med Hypotheses, 2020. **143**: p. 110087.
174. Urciuoli, E. and B. Peruzzi, *Inhibiting Extracellular Vesicle Trafficking as Antiviral Approach to Corona Virus Disease 2019 Infection*. Front Pharmacol, 2020. **11**: p. 580505.
175. Maaroufi, H., 2020.
176. Miorin, L., et al., *SARS-CoV-2 Orf6 hijacks Nup98 to block STAT nuclear import and antagonize interferon signaling*. Proc Natl Acad Sci U S A, 2020. **117** (45): p. 28344-28354.
177. Burtscher, J., et al., *Mitochondria: In the Cross Fire of SARS-CoV-2 and Immunity*. iScience, 2020. **23** (10): p. 101631.
178. Recalcati, S., et al., *Acral cutaneous lesions in the time of COVID-19*. J Eur Acad Dermatol Venereol, 2020. **34** (8): p. e346-e347.
179. Gianotti, R., et al., *Cutaneous Clinico-Pathological Findings in three COVID-19-Positive Patients Observed in the Metropolitan Area of Milan, Italy*. Acta Derm Venereol, 2020. **100** (8): p. adv00124.
180. Colmenero, I., et al., *SARS-CoV-2 endothelial infection causes COVID-19 chilblains: histopathological, immunohistochemical and ultrastructural study of seven paediatric cases*. Br J Dermatol, 2020. **183** (4): p. 729-737.
181. Lahiri, D., et al., *Neuroinvasive potential of a primary respiratory pathogen SARS- CoV2: Summarizing the evidences*. Diabetes Metab Syndr, 2020. **14** (5): p. 1053-1060.
182. Estrada, E., *Protein-driven mechanism of multiorgan damage in COVID-19*. Med Drug Discov, 2020: p. 100069.
183. Klimpel, G., *Immune Defenses*, in *Medical Microbiology. 4th edition*, Baron S, Editor. 1996.
184. Kaser, A., *Genetic risk of severe Covid-19*. New England Journal of Medicine, 2020. **383** (16): p. 1590-1591.
185. Pisanti, S., et al., *Correlation of the two most frequent HLA haplotypes in the Italian population to the differential regional incidence of Covid-19*. Journal of Translational Medicine, 2020. **18** (1).
186. Hou, Y., et al., *New insights into genetic susceptibility of COVID-19: An ACE2 and TMPRSS2 polymorphism analysis*. BMC Medicine, 2020. **18** (1).
187. Van Der Made, C.I., et al., *Presence of Genetic Variants among Young Men with Severe COVID-19*. JAMA - Journal of the American Medical Association, 2020. **324** (7): p. 663-673.
188. Gracia-Ramos, A.E., *Is the ACE2 Overexpression a Risk Factor for COVID-19 Infection?* Archives of Medical Research, 2020. **51** (4): p. 345-346.
189. Wallentin, L., et al., *Angiotensin-converting enzyme 2 (ACE2) levels in relation to risk factors for COVID-19 in two large cohorts of patients with atrial fibrillation*. European heart journal, 2020. **41** (41): p. 4037-4046.
190. Strobe, J.D., C.H.C. PharmD, and W.D. Figg, *TMPRSS2: Potential Biomarker for COVID-19 Outcomes*. Journal of Clinical Pharmacology, 2020. **60** (7): p. 801-807.
191. Miesbach, W. and M. Makris, *COVID-19: Coagulopathy, Risk of Thrombosis, and the Rationale for Anticoagulation*. Clinical and Applied Thrombosis/Hemostasis, 2020. **26**.
192. Laguna-Goya, R., et al., *IL-6-based mortality risk model for hospitalized patients with COVID-19*. Journal of Allergy and Clinical Immunology, 2020. **146** (4): p. 799-807.e9.
193. Zuo, Y., et al., *Neutrophil extracellular traps in COVID-19*. JCI Insight, 2020. **5** (11).
194. Chiappetta, S., et al., *COVID-19 and the role of chronic inflammation in patients with obesity*. International Journal of Obesity, 2020. **44** (8): p. 1790-1792.
195. Cecchini, R. and A.L. Cecchini, *SARS-CoV-2 infection pathogenesis is related to oxidative stress as a response to aggression*. Med Hypotheses, 2020. **143**: p. 110102.
196. Burtscher, J., G.P. Millet, and M. Burtscher, *Low cardiorespiratory and mitochondrial fitness as risk factors in viral infections: implications for COVID-19*. Br J Sports Med, 2020.
197. Mahase, E., *Covid-19: Why are age and obesity risk factors for serious disease?* BMJ Case Reports, 2020. **371**: p. m4130.
198. *COVID-19 Hospitalization and Death by Race/Ethnicity*. 2020, CDC.
199. Wu, M., et al., *Transcriptional and proteomic insights into the host response in fatal COVID-19 cases*. Proceedings of the National Academy of Sciences of the United States of America, 2020. **117** (45): p. 28336-28343.
200. Veenstra, J., et al., *Antecedent immunosuppressive therapy for immune-mediated inflammatory diseases in the setting of a COVID-19 outbreak*. Journal of the American Academy of Dermatology, 2020. **83** (6): p. 1696-1703.
201. Irfan, M., et al., *Pulmonary functions in patients with diabetes mellitus*. Lung India, 2011. **28** (2): p. 89-92.
202. Zhong, J., Q. Gong, and A. Mima, *Inflammatory Regulation in Diabetes and Metabolic Dysfunction*. Journal of Diabetes Research, 2017. **2017**.
203. Kulkarni, S., B.L. Jenner, and I. Wilkinson, *COVID-19 and hypertension*. JRAAS - Journal of the Renin-Angiotensin-Aldosterone System, 2020. **21** (2).

204. Shah, B., P. Modi, and S.R. Sagar, *In silico studies on therapeutic agents for COVID-19: Drug repurposing approach*. Life Sciences, 2020. **252**.
205. Ahmed, S.A., et al., *Destabilizing the structural integrity of COVID-19 by caulerpin and its derivatives along with some antiviral drugs: An in silico approaches for a combination therapy*. Structural Chemistry, 2020. **31** (6): p. 2391-2412.
206. Ge, C. and Y. He, *In Silico Prediction of Molecular Targets of Astragaloside IV for Alleviation of COVID-19 Hyperinflammation by Systems Network Pharmacology and Bioinformatic Gene Expression Analysis*. Frontiers in Pharmacology, 2020. **11**.
207. Garcia-Cremades, M., et al., *Optimizing Hydroxychloroquine Dosing for Patients With COVID-19: An Integrative Modeling Approach for Effective Drug Repurposing*. Clinical Pharmacology and Therapeutics, 2020. **108** (2): p. 253-263.
208. Wang, S., et al., *Modeling the viral dynamics of SARS-CoV-2 infection*. Math Biosci, 2020. **328**: p. 108438.
209. Taguchi, Y.H. and T. Turki, *A new advanced in silico drug discovery method for novel coronavirus (SARS-CoV-2) with tensor decomposition-based unsupervised feature extraction*. PLoS ONE, 2020. **15** (9 September).
210. Mulay, A., et al., *SARS-CoV-2 infection of primary human lung epithelium for COVID-19 modeling and drug discovery*. bioRxiv, 2020.
211. Touret, F., et al., *In vitro screening of a FDA approved chemical library reveals potential inhibitors of SARS-CoV-2 replication*. Scientific Reports, 2020. **10** (1).
212. Canham, M.A., J.D.M. Campbell, and J.C. Mountford, *The use of mesenchymal stromal cells in the treatment of coronavirus disease 2019*. J Transl Med, 2020. **18** (1): p. 359.
213. Kavianpour, M., M. Saleh, and J. Verdi, *The role of mesenchymal stromal cells in immune modulation of COVID-19: focus on cytokine storm*. Stem Cell Res Ther, 2020. **11** (1): p. 404.
214. McAuley, D.F., et al., *Clinical grade allogeneic human mesenchymal stem cells restore alveolar fluid clearance in human lungs rejected for transplantation*. Am J Physiol Lung Cell Mol Physiol, 2014. **306** (9): p. L809-15.
215. Al-Khawaga, S. and E.M. Abdelalim, *Potential application of mesenchymal stem cells and their exosomes in lung injury: an emerging therapeutic option for COVID-19 patients*. Stem Cell Res Ther, 2020. **11** (1): p. 437.
216. Han, Y., et al., *Identification of Candidate COVID-19 Therapeutics using hPSC-derived Lung Organoids*. bioRxiv, 2020.
217. Han, Y., et al., *Identification of SARS-CoV-2 inhibitors using lung and colonic organoids*. Nature, 2020.
218. Si, L., et al., 2020.
219. Bassett, C.A., *Beneficial effects of electromagnetic fields*. J Cell Biochem, 1993. **51** (4): p. 387-93.
220. Premi, E., et al., *Modulation of long-term potentiation-like cortical plasticity in the healthy brain with low frequency-pulsed electromagnetic fields*. BMC Neurosci, 2018. **19** (1): p. 34.
221. MITCHELL, H.H., HAMILTON, T. S., STEGGERDA, li'. R., BEAN, H. W., *THE CHEMICAL COMPOSITION OF THE ADULT HUMAN BODY AND ITS BEARING ON THE BIOCHEMISTRY OF GROWTH*. JBC, 1945. **158**: p. 625-637.
222. Vigano, M., et al., *Mesenchymal stem cells as therapeutic target of biophysical stimulation for the treatment of musculoskeletal disorders*. J Orthop Surg Res, 2016. **11** (1): p. 163.
223. Chan, A.K., et al., *Pulsed electromagnetic fields reduce acute inflammation in the injured rat-tail intervertebral disc*. JOR Spine, 2019. **2** (4): p. e1069.
224. de Girolamo, L., et al., *In vitro functional response of human tendon cells to different dosages of low-frequency pulsed electromagnetic field*. Knee Surg Sports Traumatol Arthrosc, 2015. **23** (11): p. 3443-53.
225. Saraiva, M. and A. O'Garra, *The regulation of IL-10 production by immune cells*. Nat Rev Immunol, 2010. **10** (3): p. 170-81.
226. Akahoshi, T., J.J. Oppenheim, and K. Matsushima, *Interleukin 1 stimulates its own receptor expression on human fibroblasts through the endogenous production of prostaglandin(s)*. J Clin Invest, 1988. **82** (4): p. 1219-24.
227. Petersen, A.M. and B.K. Pedersen, *The anti-inflammatory effect of exercise*. J Appl Physiol (1985), 2005. **98** (4): p. 1154-62.
228. JE., d.V., *Immunosuppressive and anti-inflammatory properties of interleukin 10*. Ann Med., 1995. **27** (5): p. 537-41.
229. al., J.B.M.e., *IL-10 Elicits IFN γ -Dependent Tumor Immune Surveillance*. Cancer Cell, 2011. **20** (6): p. 781-96.
230. Bonnans C, C.J., Werb Z., *Remodelling the extracellular matrix in development and disease*. Nat Rev Mol Cell Biol., 2014. **15** (12): p. 786-801.
231. Pietramaggiore G, L.P., Scherer SS, Kaipainen A., *Tensile forces stimulate vascular remodeling and epidermal cell proliferation in living skin*. Ann Surg 2007. **246**: p. 896.
232. Lieleg O, B.R., Bausch AR., *Selective filtering of particles by the extracellular matrix: an electrostatic bandpass*. Biophys J. , 2009. **16** (97(6)): p. 1569-77.
233. DE, I., *Cellular mechanotransduction: putting all the pieces together again*. FASEB Journal. **20**: p. 811.
234. al., B.E.e., *Th17: the third member of the effector T cell trilogy*. Current Opinion in Immunology, 2007. **19**: p. 652-657.
235. Yalcinkaya E, C.M., Bugan B. , *Extracellular matrix turnover: a balance between MMPs and their inhibitors*. Arq Bras Cardiol. , 2014. **102** (5): p. 519-20.
236. Siwik DA, C.W., *Regulation of matrix metalloproteinases by cytokines and reactive oxygen/nitrogen species in the myocardium*. Heart Fail Rev., 2004. **9** (1): p. 43-51.
237. LJ., Y., *Positive oxidative stress in aging and aging-related disease tolerance*. Redox Biol., 2014. **2**: p. 165-9.
238. al., S.P.e., *Interleukin (IL)-6 and Its Soluble Receptor Induce TIMP-1 Expression in Synoviocytes and Chondrocytes, and Block IL-1-induced Collagenolytic Activity*. J Biol Chem. , 1998. **273**: p. 13625-13629.
239. al., S.A.e., *Interleukin 8 (monocyte-derived neutrophil chemotactic factor) dynamically regulates its own receptor expression on human neutrophils*. J Biol Chem., 1990. **265** (1): p. 183-9.
240. Glocker EO, e.a., *IL-10 and IL-10 receptor defects in humans*. Ann N Y Acad Sc, 2011. **1246**: p. 102-7.
241. Zhang W, e.a., *Protective effect of bone marrow mesenchymal stem cells in intestinal barrier permeability after heterotopic intestinal transplantation*. World J Gastroenterol., 2014. **20** (23): p. 7442-51.
242. Garcia-Hernández V, e.a., *EGF regulates claudin-2 and -4 expression through Src and STAT3 in MDCK cells*. J Cell Physiol., 2015. **230** (1): p. 105-15
243. Howe KL, e.a., *Transforming growth factor-beta regulation of epithelial tight junction proteins enhances barrier function and blocks enterohemorrhagic Escherichia coli O157:H7-induced increased permeability*. Am J Pathol., 2005. **167** (6): p. 1587-97.
244. Xiong, J., et al., *Effects of interleukin-4 or interleukin-10 gene therapy on trinitrobenzenesulfonic acid-induced murine colitis*. BMC Gastroenterol, 2013. **13**: p. 165.
245. Bao, S. and D.L. Knoell, *Zinc modulates cytokine-induced lung epithelial cell barrier permeability*. Am J Physiol Lung Cell Mol Physiol, 2006. **291** (6): p. L1132-41.
246. Liu, M.J., et al., *Zinc regulates the acute phase response and serum amyloid A production in response to sepsis through JAK-STAT3 signaling*. PLoS One, 2014. **9** (4): p. e94934.
247. Chen, Y., et al., *Vitamin D receptor inhibits nuclear factor kappaB activation by interacting with IkappaB kinase beta protein*. J Biol Chem, 2013. **288** (27): p. 19450-8.
248. Jeffery, L.E., et al., *1,25-Dihydroxyvitamin D3 and IL-2 combine to inhibit T cell production of inflammatory cytokines and promote development of regulatory T cells expressing CTLA-4 and FoxP3*. J Immunol, 2009. **183** (9): p. 5458-67.
249. Ellulu, M.S., et al., *Effect of vitamin C on inflammation and metabolic markers in hypertensive and/or diabetic obese adults: a randomized controlled trial*. Drug Des Devel Ther, 2015. **9**: p. 3405-12.
250. Derosa, G., et al., *A role for quercetin in coronavirus disease 2019 (COVID-19)*. Phytother Res, 2020.
251. Yang, M., et al., *Resveratrol inhibits the replication of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) in cultured Vero cells*. Phytother Res, 2020.
252. Das, S., et al., *An investigation into the identification of potential inhibitors of SARS-CoV-2 main protease using molecular docking study*. J Biomol Struct Dyn, 2020: p. 1-11.

253. Li, H.Y., et al., *Curcumin inhibits angiotensin II-induced inflammation and proliferation of rat vascular smooth muscle cells by elevating PPAR-gamma activity and reducing oxidative stress*. *Int J Mol Med*, 2017. **29** (5): p. 1307-1316.
254. Garcia-Mauriño S, G.-H.M., Calvo JR, Rafii-El-Idrissi M, Sanchez-Margalet V, Goberna R, Guerrero JM., *Melatonin enhances IL-2, IL-6, and IFN-gamma production by human circulating CD4+ cells: a possible nuclear receptor-mediated mechanism involving T helper type 1 lymphocytes and monocytes*. *J Immunol.*, 1997. **15** (2): p. 574-81.
255. Bellavite, P. and A. Donzelli, *Hesperidin and SARS-CoV-2: New Light on the Healthy Function of Citrus Fruits*. *Antioxidants (Basel)*, 2020. **9** (8).
256. Lau, J.T., et al., *The use of an herbal formula by hospital care workers during the severe acute respiratory syndrome epidemic in Hong Kong to prevent severe acute respiratory syndrome transmission, relieve influenza-related symptoms, and improve quality of life: a prospective cohort study*. *J Altern Complement Med*, 2005. **11** (1): p. 49-55.
257. Huang, F., et al., *A review of therapeutic agents and Chinese herbal medicines against SARS-COV-2 (COVID-19)*. *Pharmacol Res*, 2020. **158**: p. 104929.
258. Panyod, S., C.T. Ho, and L.Y. Sheen, *Dietary therapy and herbal medicine for COVID-19 prevention: A review and perspective*. *J Tradit Complement Med*, 2020. **10** (4): p. 420-427.
259. Serafino, A., et al., *Stimulatory effect of Eucalyptus essential oil on innate cell-mediated immune response*. *BMC Immunol*, 2008. **9**: p. 17.